SYNTHESIS AND PHARMACOLOGICAL CHARACTERIZATION OF CERTAIN BACLOFEN ANALOGUES

MOHAMED I. ATTIA^{a,b*}, CLAUS HERDEIS^{b*}, HANS BRÄUNER-OSBORNE^c ^aDepartment of Pharmaceutical Chemistry, College of Pharmacy, King Saud University, Riyadh 11451, Saudi Arabia

^bDepartment of Pharmaceutical Chemistry, Institute of Pharmacy and Food Chemistry, Würzburg University, Am Hubland, 97074, Würzburg, Germany ^cDepartment of Medicinal Chemistry, Faculty of Pharmaceutical Sciences, University of Copenhagen, Universitetsparken 2, DK-2100 Copenhagen, Denmark

(*RS*)-4-Amino-3-(4-chlorophenyl)butanoic acid (baclofen, **2**) is the prototypic selective GABA_BR agonist and is used clinically for the treatment of spasticity associated with brain and spinal cord injuries. Synthesis and GABA_B receptor agonist activity of certain amino acids structurally related to baclofen (**2**) are reported. (*RS*)-4-amino-3-(4-ethynylphenyl)butanoic acid hydrochloride (**1b**) showed GABA_B receptor agonist activity with $EC_{50} = 240 \ \mu M$ whereas (*RS*)-4-amino-3-(4-iodophenyl)butanoic acid hydrochloride (**1c**) emerged as the best GABA_B receptor agonist congener in the synthesized compounds with $ED_{50} = 32 \ \mu M$.

(Received November 14, 2012; Accepted December 20, 2012)

Keywords: GABA; Synthesis; Baclofen analogues; GABA_B receptor agonists; Pharmacological characterization.

1. Introduction

The principle inhibitory neurotransmitter in the central nervous system, γ -aminobutyric acid (GABA, **1**, Figure 1), exerts its effect through three different receptor subtypes: ligand-gated chloride channels GABA_A and GABA_C receptors, and G-protein-coupled metabotropic (GPCRs) GABA_B receptors [1]. GABA_B receptors were discovered in 1980 by Norman G. Bowery [2] and act post- and pre-synaptically to inhibit neuronal excitability and neurotransmitter release, respectively. The molecular structure of GABA_B receptor was cloned in 1997 [3]. GABA_B receptor is composed of two subunits, GABA_{B1} and GABA_{B2}, to form a heterodimer which is necessary for GABA_B receptors to be functionally active [4]. The former subunit contains the GABA binding domain, while GABA_{B2} subunit provides the G-protein-coupling mechanism and also incorporates an allosteric modulatory site within its heptahelical structure [5].

(*RS*)-4-Amino-3-(4-chlorophenyl)butanoic acid (baclofen, **2**, Figure 1), the aromatic analogue of GABA, introduced as racemate in 1973 in the therapy of muscle spasticity, is still the classical prototype of the selective GABA_BR agonists [6]. The most predominant pharmacological effects of baclofen are the muscle relaxant, anti-nociceptive and anti-drug craving effects, as well as the reduction in cognitive behavior [7]. (*RS*)-Baclofen (**2**) is used clinically for the treatment of spasticity associated with brain and spinal cord injuries [8], drug addiction and alcoholism [9], gastroesophageal reflux disease (GERD) [10], cancer pain [11] and overactive bladder [12]. The GABA_B activity of racemic baclofen is known to reside primarily in the *R*-(-)-enantiomer [13].

^{*}Corresponding author: mattia@ksu.edu.sa

Recently, *R*-(-)-baclofen is under development for the treatment of behavioral symptoms of Fragile X Disorder [6].

On the other hand, δ -aminovaleric acid (DAVA, **3**, Figure 1), the homologue of GABA, is a nonselective GABA ergic receptor antagonist [14].



Fig. 1: Chemical structures of GABA, Baclofen and DAVA.

Relatively few investigations have been documented on baclofen analogues having different aromatic substituents since the initial synthesis of baclofen more than 40 years ago [15]. Accordingly, we report herein the synthesis and GABA_B agonist activity of compound **1a** with an integrated aromatic ring in the backbone of DAVA (**3**), as well as the ethynyl and iodo analogues of baclofen, namely compounds **1b**, and **1c**, respectively (Figure 2).



Fig. 2: Chemical structures of the target compounds 1a-c.

2. Experimental

2.1. Chemistry

Melting points were determined using a capillary melting point apparatus (Gallenkamp, Sanyo) and are uncorrected. Infrared (IR) spectra were recorded as thin layer films (for oils) or as pellets (for solids) with BIO-RAD spectrometer and values are represented in cm⁻¹. NMR (¹H NMR and ¹³C NMR) spectra were carried out on AC 250 Bruker spectrometer (250 MHz for ¹H NMR and 63 MHz for ¹³C NMR) and chemical shift values were recorded in ppm on δ scales. All samples were measured at room temperature. The ¹H NMR data are represented as follows: chemical shifts, multiplicity and number of protons. Column chromatography was carried out on silica gel 60 (0.063–0.200 mm) obtained from Merck. Elemental analyses were performed by the microanalytical section of the Institute of Inorganic Chemistry, University of Würzburg.

2.1.1. Synthesis of 5-chloroisophthalic acid (8a)

Sodium nitrite (2.10 g, 30 mmol) was added portion wise to a stirred concentrated sulfuric acid (22 ml) over a period of 10-15 min at ambient temperature. After the addition was completed, the temperature was raised to 70 °C and the mixture was stirred until all the sodium nitrite was dissolved. The resulting solution was added dropwise to a cold (5-10 °C) stirred suspension of 5-aminoisophthalic acid (**9a**) (5 g, 28 mmol) in glacial acetic acid (60 ml). After the addition was completed, the solution was stirred at 40 °C for 30 min. The resulting diazonium salt was added in

140

portions over a period of 5 min to a cold (0-5 °C) stirred solution of cuprous chloride (5.60 g, 57 mmol) in concentrated hydrochloric acid (60 ml). After complete addition, the mixture was heated to 80 °C under stirring for 20 min. An equal volume of water was then added and the mixture was cooled in an ice bath. After 2 h, the white crystals were filtered, washed with water

and dried under vacuum to give 4.54 g (82 %) of **8a** as a white crystals, m.p. 278-280 °C (lit.[16] 277 °C). This material was pure enough to be used in the next step without further purification.

IR (neat, cm⁻¹) exhibited bands at 3982-2523, 1693, 1605, 1574, 1400, 1271. ¹H NMR (DMSO- d_6): δ (ppm) = 8.06 (d, J = 1.53 Hz, 2H, H_{arom}), 8.35 (t, J = 1.53 Hz, 1H, H_{arom}). ¹³C NMR (DMSO- d_6): δ (ppm) = 129.2, 133.6, 134.1, 134.6 (C_{arom}), 166.2 (carbonyl acid).

2.1.2. Synthesis of 5-chloroisophthalic acid dimethyl ester (7a)

A mixture of 5-chloroisophthalic acid (**8a**) (2.05 g, 10 mmol) and concentrated sulfuric acid (0.52 ml) in methanol (40 ml) was refluxed with stirring for 24 h. The reaction mixture was evaporated under reduced pressure, the residue was dissolved in dichloromethane (25 ml) and washed with saturated sodium bicarbonate solution (2 x 15 ml). The organic layer was dried (Na₂SO₄), evaporated under vacuum to give 2.15 g (92 %) of **7a** [17] as a white powder m.p. 76-78 °C which was used in the next step without further purification. IR (neat, cm⁻¹) exhibited bands at 1731, 1582, 1431, 1312, 1248. ¹H NMR (DMSO-*d*₆): δ (ppm) = 3.90 (s, 6H, 2 x OCH₃), 8.04 (d, *J* = 1.23 Hz, 2H, H_{arom}), 8.24 (t, *J* = 1.23 Hz, 1H, H_{arom}). ¹³C NMR (DMSO-*d*₆): δ (ppm) = 53.3 (OCH₃), 128.7, 132.8, 133.7, 134.9 (C_{arom}), 164.9 (ester carbonyl).

2.1.3. Synthesis of 5-chloroisophthalic acid monomethyl ester (6a)

1 N NaOH (8.00 ml, 8 mmol) was added to a solution of 5-chloroisophthalic acid dimethyl ester (**7a**) (2.06 g, 9 mmol) in methanol (75 ml). The reaction mixture was stirred at ambient temperature for 18 h. The solvent was removed under reduced pressure, the residue was dissolved in saturated sodium bicarbonate solution (25 ml) and extracted with dichloromethane (2 x 10 ml). The aqueous layer was cooled (5-10 °C), acidified using concentrated hydrochloric acid and extracted with dichloromethane (3 x 20 ml). The combined organic layers were dried (Na₂SO₄), evaporated under vacuum to give 1.49 g (77 %) of **6a** [17] as a white powder m.p. 172-174 °C which was pure enough to be used in the next step. IR (neat, cm⁻¹) exhibited bands at 3085-2539, 1736, 1703, 1602, 1580, 1309, 1262. ¹H NMR (DMSO-*d*₆): δ (ppm) = 3.91 (s, 3H, OCH₃), 8.04-8.09 (m, 2H, H_{arom}), 8.32 (t, *J* = 1.23 Hz, 1H, H_{arom}). ¹³C NMR (DMSO-*d*₆): δ (ppm) = 53.6 (OCH₃), 128.9, 132.8, 133.3, 133.9, 134.2, 134.8 (carom.), 165.1 (ester carbonyl), 165.9 (carboxylic acid).

2.1.4. Synthesis of 3-chloro-5-hydroxymethylbenzoic acid methyl ester (5a) To a solution of 5-chloroisophthalic acid monomethyl ester (6a) (1.07 g, 5 mmol) in tetrahydrofuran (15 ml) was added dropwise borane-dimethyl sulfide complex (1.01 ml, 10.0 mmol). The reaction mixture was stirred at ambient temperature for 30 h. Methanol (20 ml) was carefully added dropwise to the reaction mixture and stirring was continued for further 30 min. The reaction mixture was evaporated under reduced pressure, the residue was dissolved in dichloromethane (15 ml) and washed with saturated sodium bicarbonate solution (2 x 10 ml). The organic layer was dried (Na₂SO₄), evaporated under vacuum to afford 0.87g (87 %) of **5a** [17] as a pale yellow oil which was used in the next step without further purification. IR (neat, cm⁻¹) exhibited bands at 3600-3150, 1719, 1582, 1433, 1285. ¹H NMR (CDCl₃): δ (ppm) = 2.88 (br. s, 1H, OH), 3.69 (s, 3H, OCH₃), 4.46 (s, 2H, CH₂OH), 7.29 (s, 1H, H_{arom}), 7.61 (s, 1H, H_{arom}), 7.64 (t, *J* = 1.53 Hz, 1H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 52.9 (OCH₃), 64.2 (CH₂OH), 126.2, 128.9 , 131.5, 132.1, 135.0, 143.7 (C_{arom}), 166.4 (ester carbonyl).

2.1.5. Synthesis of 3-bromomethyl-5-chlorobenzoic acid methyl ester (4a)

Phosphorous tribromide (0.87 ml, 9 mmol) was added dropwise to a cold (-30 °C), stirred 3-chloro-5-hydroxymethylbenzoic acid methyl ester (**5a**) (0.60 g, 3 mmol). The reaction mixture was allowed to warm to room temperature and stirred for further 30 min at ambient temperature. The reaction mixture was carefully poured into ice-cold water (20 ml) and extracted with diethyl

142

ether (3 x 15 ml). The combined ether extracts were dried (Na₂SO₄), evaporated under reduced pressure and the residue was purified by column chromatography using petroleum ether (40-60 °C): diethyl ether (8 : 2) to give 0.47 g (60 %) of **4a** [17] as a white powder m.p. 56-58 °C. IR (neat, cm⁻¹) exhibited bands at 1717, 1578, 1448, 1429, 1291. ¹H NMR (CDCl₃) δ (ppm) = 3.96 (s, 3H, OCH₃), 4.48 (s, 2H, CH₂Br), 7.59 (t, *J* = 1.83 Hz, 1H, H_{arom}), 7.96 (d, *J* = 1.83 Hz, 2H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 31.6 (CH₂Br), 52.9 (OCH₃), 128.7, 129.9, 132.7, 133.7, 135.2, 140.4 (Carom), 165.7 (ester carbonyl).

2.1.6. Synthesis of 3-azidomethyl-5-chlorobenzoic acid methyl ester (3a)

Sodium azide (1.30 g, 20 mmol) was added to a solution of 3-bromomethyl-5chlorobenzoic acid methyl ester (**4a**) (0.79 g, 3 mmol) in acetone (15 ml) and water (4 ml). The reaction mixture was refluxed for 18 h. The solvent was removed under reduced pressure, the residue was dissolved in dichloromethane (20 ml) and washed with water (3 x 10 ml). The organic layer was dried (Na₂SO₄) and evaporated under vacuum. The residue was purified by column chromatography using petroleum ether (40-60 °C): diethyl ether (8 : 2) to afford 0.58 g (86%) of **3a** as a pale yellow oil. IR (neat, cm⁻¹) exhibited bands at 2098, 1723, 1582, 1433, 1283. ¹H NMR (CDCl₃): δ (ppm) = 3.96 (s, 3H, OCH₃), 4.43 (s, 2H, -CH₂-N=), 7.53 (t, *J* = 1.53 Hz, 1H, H_{arom}), 7.89 (s, 1H, H_{arom}), 7.99 (t, *J* = 1.53 Hz, 1H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 52.9 (OCH₃), 54.1 (-CH₂-N=), 127.6, 129.8, 132.6, 132.7, 135.4, 138.3 (C_{arom}), 165.8 (ester carbonyl).

2.1.7. Synthesis of 3-azidomethyl-5-chlorobenzoic acid (2a)

1 N NaOH (2.00 ml, 2 mmol) was added to a solution of 3- azidomethyl–5-chlorobenzoic acid methyl ester (**3a**) (0.23 g, 1 mmol) in methanol (15 ml). The reaction mixture was refluxed for 2.5 h. The solvent was removed under reduced pressure, the residue was dissolved in saturated sodium bicarbonate solution (10 ml) and extracted with dichloromethane (2 x 5 ml). The aqueous layer was cooled (0-5 °C), acidified with concentrated hydrochloric acid and extracted with dichloromethane (3 x 10 ml). The combined organic extracts were dried (Na₂SO₄) and evaporated under vacuum to give 0.18 g (83%) of **2a** as a pale yellow powder m.p. 92-94 °C and was used in the next step without further purification. IR (neat, cm⁻¹) exhibited bands at 3093-2406, 2097, 1687, 1603, 1583, 1410, 1295. ¹H NMR (CDCl₃): δ (ppm) = 4.47 (s, 2H, -CH₂-N=), 7.59 (t, *J* = 1.83 Hz, 1H, H_{arom}), 7.96 (s, 1H, H_{arom}, 8.06 (t, *J* = 1.83 Hz, 1H, H_{arom}), 11.8 (br.s, 1H, COOH). ¹³C NMR (CDCl₃): δ (ppm) = 54.0 (-CH₂-N=), 128.1, 130.4, 131.8, 133.6, 135.6, 138.5 (C_{arom}), 171.2 (carboxylic acid). C₈H₆ClN₃O₂: calcd. C 45.41, H 2.86, N 19.86; found C 45.33, H 2.95, N 19.46.

2.1.8. Synthesis of 3-aminomethyl-5-chlorobenzoic acid hydrochloride (1a)

To a solution of 3-azidomethyl-5-chlorobenzoic acid (**2a**) (0.21 g, 1 mmol) in 95% 2propanol (10 ml) and concentrated hydrochloric acid (0.50 ml) was added PtO₂ (0.04 g). The mixture was hydrogenated on a Parr shaker apparatus under 4 bar pressure of hydrogen gas for 18 h at ambient temperature. The catalyst was removed by filtration and the solvent was evaporated under vacuum. The residue was dissolved in water (10 ml) and extracted with dichloromethane (2 x 25 ml). The aqueous layer was evaporated under reduced pressure and the residue was recrystallized (2-propanol /diethyl ether) to afford 0.18 g (82 %) of **1a** as a white hygroscopic powder m.p. 260-263 °C. IR (neat, cm⁻¹) exhibited bands at 3137-2887, 1709, 1607, 1581, 1395, 1207. ¹H NMR (D₂O): δ (ppm) = 4.13 (s, 2H, -CH₂-NH₂), 7.59 (t, *J* = 1.83 Hz, 1H, H_{arom}.), 7.81 (t, *J* = 1.83 Hz, 2H, H_{arom}). ¹³C NMR (D₂O): δ (ppm) = 42.6 (-CH₂-NH₂), 128.6, 130.5, 132.9, 133.9, 135.0, 135.2 (C_{arom}), 168.9 (carboxylic acid). C₈H₉Cl₂NO₂: calcd. C 43.27, H 4.09, N 6.31; found C 43.49, H 4.23, N 6.28.

2.1.9. Synthesis of (E)-3-(4-iodophenyl)acrylic acid methyl ester (7b)

To a solution of (methoxycarbonylmethylene)triphenylphosphorane [18] (3.35 g, 10 mmol) in dry dichloromethane (10 ml) was added dropwise a solution of 4-iodobenzaldehyde (**8b**) (2.32 g, 10 mmol) in dichloromethane (5 ml). The resulting pale yellow solution was stirred at room temperature for one hour (NMR monitoring). The reaction mixture was evaporated under vacuum and the residue was purified by column chromatography using petroleum ether (40-60

°C): diethyl ether (9 : 1) to yield 2.68 g (93%) of **7b** [18] as a pale yellow powder m.p. 128-130 °C. IR (neat, cm⁻¹) exhibited bands at 1711, 1636, 1437, 1310, 1189. ¹H NMR (CDCl₃): δ (ppm) = 3.83 (s, 3H, OCH₃), 6.74 (d, *J* = 16.18 Hz, 1H, 2-H), 7.27 (d, *J*_{AB} = 8.53 Hz, 2H, H_{arom}), 7.63 (d, *J* = 16.15 Hz, 1H, 3-H), 7.75 (d, *J*_{AB} = 8.55 Hz, 2 H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 52.2 (OCH₃), 96.9 (C_{arom}), 118.9 (C-2), 129.9, 134.3, 138.5 (C_{arom}), 144.0 (C-3), 167.5 (C-1). C₁₀H₉IO₂: calcd. C 41.69, H 3.15; found C 41.29, H 3.15.

2.1.10. Synthesis of (RS)-3-(4-iodophenyl)-4-nitrobutanoic acid methyl ester (6b)

To a stirred solution of (*E*)-3-(4-iodophenyl)acrylic acid methyl ester (**7b**) (2 g, 7 mmol) in nitromethane (14 ml) was added Triton B (0.6 ml). The resulting pale yellow solution was heated at 85 °C for two hours. The reaction mixture was cooled (0-5 °C), acidified using 1 N hydrochloric acid and evaporated under reduced pressure. The residue was dissolved in diethyl ether (20 ml) and washed with water (2 x 10 ml). The organic layer was dried (Na₂SO₄), evaporated under vacuum and the residue was purified by column chromatography using petroleum ether (40-60 °C): diethyl ether (1 : 1) to give 2.18 g (89%) of **6b** as a white powder m.p. 58-60 °C. IR (neat, cm⁻¹) exhibited bands at 1730, 1721, 1561, 1536, 1374. ¹H NMR (CDCl₃): δ (ppm) = 2.77-2.85 (m, 2H, 2-H), 3.66 (s, 3H, OCH₃), 3.96-4.02 (m, 1H, 3-H), 4.63 (dd, *J*_{4a-3} = 8.25 Hz, *J*_{gem} = 12.83 Hz, 1H, 4-H_a), 4.74 (dd, *J*_{4b-3} = 6.7 Hz, *J*_{gem} = 12.83 Hz, 1H, 4-H_b), 7.01 (d, *J*_{AB} = 8.25 Hz, 2H, H_{arom}), 7.69 (d, *J*_{AB} = 8.53 Hz, 2H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 37.7 (C-2), 40.1 (C-3), 52.5 (OCH₃), 79.4 (C-4), 94.1, 129.7, 138.4, 138.6 (C_{arom}), 171.2 (C-1). C₁₁H₁₂INO₄: calcd. C 37.84, H 3.46, N 4.01; found C 37.80, H 3.38, N 3.71.

2.1.11. Synthesis of (RS)-4-(4-iodophenyl)-2-pyrrolidinone (5b)

To a cold (0 °C) mixture of concentrated hydrochloric acid (1.5 ml) and methanol (1.5 ml) was added simultaneously in portions both zinc dust (0.46 g, 7 mmol) and a solution of (*RS*)-3-(4-iodophenyl)-4-nitrobutanoic acid methyl ester (**6b**) (0.25 g, 0.7 mmol) in methanol (3 ml) through 30 min. The reaction mixture was further stirred for 30 min at 0 °C. The reaction mixture was filtered and the filtrate was evaporated under reduced pressure. The residue was dissolved in water (10 ml), extracted with diethyl ether (3 x 5 ml). The aqueous layer was basified with 6 N sodium hydroxide solution (15 ml), extracted with diethyl ether (3 x 20 ml). The combined organic extracts were dried (Na₂SO₄) and evaporated under vacuum to afford 0.13 g (65%) of **5b** as a white powder m.p. 144-145 °C (lit. [20] 143-145 °C) which was pure enough to be used in the next step without further purification. IR (neat, cm⁻¹) exhibited bands at 3181, 1703, 1477, 1285, 1267. ¹H NMR (CDCl₃): δ (ppm) = 2.28 (dd, $J_{3a-4} = 8.55$ Hz, $J_{gem} = 16.78$ Hz, 1H, 3-Ha), 2.77 (dd, $J_{3b-4} = 8.85$ Hz, $J_{gem} = 16.78$ Hz, 1H, 3-Hb), 3.41 (dd, $J_{5a-4} = 7$ Hz, $J_{gem} = 9.15$ Hz, 1H, 5-Ha), 3.66 (quin, $J_{4-3a} = J_{4-5b} = J_{4-5b} = 8.23$ Hz, 1H, 4-H), 3.82 (m, 1H, 5-Hb), 7.03 (d, $J_{AB} = 8.23$ Hz, 2H, H_{arom}), 7.09 (br.s, 1H, N–H), 7.68 (d, $J_{AB} = 8.23$ Hz, 2H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 38.3 (C-3), 40.2 (C-4), 49.9 (C-5), 92.7, 129.2, 138.3, 142.2 (C_{arom}), 178.1 (lactam).

2.1.12. Synthesis of (RS)-1-tert-butyloxycarbonyl-4-(4-iodophenyl)-2-pyrrolidinone (4b)

To a stirred solution of (*RS*)-4-(4-iodophenyl)-2-pyrrolidinone (**5b**) (0.23 g, 0.8 mmol), di*tert*- butyl dicarbonate (0.35 g, 1.6 mmol) and 4-(dimethylamino)pyridine (0.1 g, 0.8 mmol) in dichloromethane (15 ml) was added triethylamine (0.08 g, 0.11 ml, 0.8 mmol) under nitrogen atmosphere. The resulting solution was stirred for 30 h at ambient temperature. The volatiles were removed under reduced pressure, the residue was dissolved in diethyl ether (15 ml), washed with 1 N hydrochloric acid (2 x 5 ml), washed with water (2 x 5 ml), dried (Na₂SO₄) and evaporated under vacuum. The residue was purified by column chromatography using petroleum ether (40-60 °C): diethyl ether (1 : 1) to yield 0.26 g (84%) of **4b** as a white powder m.p. 100-102 °C. IR (neat, cm⁻¹) exhibited bands at 1774, 1696, 1369, 1343, 1288. ¹H NMR (CDCl₃): δ (ppm) = 1.31 (s, 9H, *t*-Bu.), 2.43 (dd, $J_{3a-4} = 9.45$ Hz, $J_{gem} = 17.08$ Hz, 1H, 3-H_a), 2.67 (dd, $J_{3b-4} = 8.25$ Hz, $J_{gem} = 17.08$ Hz, 1H, 3-H_b), 3.27 (quin, $J_{4.3a} = J_{4.3b} = J_{4.5a} = J_{4.5b} = 8.55$ Hz, 1H, 4-H), 3.43 (dd, $J_{5a-4} = 8.23$ Hz, $J_{gem} = 10.68$ Hz, 1H, 5-H_a), 3.93 (dd, $J_{5b-4} = 7.95$ Hz, $J_{gem} = 10.68$ Hz, 1H, 5-H_b), 6.78 (d, $J_{AB} = 8.25$ Hz, 2H, H_{arom}), 7.46 (d, $J_{AB} = 8.25$ Hz, 2H, H_{arom}). ¹³C NMR (CDCl₃): δ (ppm) = 28.4 (OC(CH₃)₃), 36.4 (C-4), 40.5 (C-3), 53.2 (C-5), 83.6 (OC(CH₃)₃), 93.1, 129.2, 138.4, 140.8

(C_{arom}), 150.2 (urethane), 172.9 (lactam). C₁₅H₁₈INO₃: calcd. C 46.53, H 4.69, N 3.62; found C 46.72, H 4.69, N 3.24.

2.1.13. Synthesis of (RS)-1-tert-butyloxycarbonyl-4-(4-trimethylsilanylethynylphenyl)-2pyrrolidinone (3b)

To a stirred suspension of (*RS*)-1-*tert*-butyloxycarbonyl-4-(4-iodophenyl)-2-pyrrolidinone (**4b**) (0.78 g, 2 mmol), bis (triphenylphosphine)palladium dichloride (0.014 g, 0.02 mmol) and cuprous iodide (0.008 g, 0.04 mmol) in triethylamine (20 ml) was added trimethylsilylacetylene (0.24 g, 0.34 ml, 2.4 mmol) under nitrogen atmosphere. The yellow reaction mixture was stirred at ambient temperature for 12 h. The black reaction mixture was filtered and washed with diethyl ether (10 ml). The filtrate was evaporated under reduced pressure, the residue was dissolved in diethyl ether (20 ml), washed with water (3 x 10 ml), dried (Na₂SO₄) and evaporated under reduced pressure. The residue was purified by column chromatography using petroleum ether (40-60 °C): diethyl ether (1 : 1) to afford 0.64 (90%) of **3b** as a pale yellow powder m.p. 118-120 °C. IR (neat, cm⁻¹) exhibited bands at 1790, 1690, 1352, 1293, 1156. ¹H NMR (CDCl₃: δ (ppm) = 0.01 (s, 9H, Si(CH₃)₃), 1.30 (s, 9H, *t*-Bu.), 2.44 (dd, *J*_{3a-4} = 9.45 Hz, *J*_{gem} = 17.38 Hz, 1H, 3-H_a), 2.66 (dd, *J*_{3b-4} = 8.25 Hz, *J*_{gem} = 17.38 Hz, 1H, 3-H_b), 3.29 (quin, *J*_{4-3a} = *J*_{4-3b} = *J*_{4-5b} = 8.53 Hz, 1H, 4-H), 3.43 (dd, *J*_{5a-4} = 8.55 Hz, *J*_{gem} = 10.68 Hz, 1H, 5- H_a), 3.92 (dd, *J*_{5b-4} = 7.93 Hz, *J*_{gem} = 10.68 Hz, 1H, 5- H_a), 3.92 (dd, *J*_{5b-4} = 7.93 Hz, *J*_{gem} = 10.68 Hz, 1H, 5- H_a), 3.92 (dd, *J*_{5b-4} = 7.93 Hz, *J*_{gem} = 10.68 Hz, 1H, 5- H_a), 3.92 (dd, *J*_{5b-4} = 7.93 Hz, *J*_{gem} = 10.68 Hz, 1H, 5- H_a), 3.63 (C-4), 40.1 (C-3), 52.9 (C-5), 83.2 (OC(CH₃)₃), 94.8, 104.5 (C≡C), 122.4, 126.7, 132.6, 141.0 (C_{arom}), 149.9 (urethane), 172.7 (lactam). C₂₀H₂₇SINO₃: calcd. C 67.19, H 7.61, N 3.92; found C 67.48, H 7.69, N 3.77.

2.1.14. Synthesis of (RS)-4-amino-3-(4-ethynylphenyl)butanoic acid hydrochloride (1b)

To a stirred solution of (RS)-1-tert-butyloxycarbonyl-4-(4-trimethylsilanylethynylphenyl)-2-pyrrolidinone (3b) (0.25 g, 0.7 mmol) in tetrahydrofuran (9 ml) was added 1 M lithium hydroxide (3 ml). The resulting reaction mixture was stirred at room temperature for 18 h. The solvent was evaporated under reduced pressure, the residue was diluted with water (10 ml) and extracted with diethyl ether (3 x 5 ml). The aqueous layer was acidified using 0.5 M potassium hydrogen sulfate solution and extracted with diethyl ether (3 x 15 ml). The organic extracts were dried (Na₂SO₄) and evaporated under vacuum to give 0.19 g (90%) of (RS)-4-tertbutyloxycarbonylamino-3-(4-ethynylphenyl)butanoic acid (2b) as a yellow powder m.p. 126-128 °C. The crude **2b** was dissolved in ~ 2.5 M dry hydrogen chloride/ethyl acetate solution (1.6 ml) and the reaction mixture was stirred at room temperature for one hour. The reaction mixture was filtered and washed with diethyl ether (5 ml). The precipitated solid was recrystallized from 2propanol/diethyl ether to yield 0.13 g (87%) of **1b** as a white powder m.p. 209-211 °C. IR (neat, cm⁻¹) exhibited bands at 3244-2654, 1724, 1520, 1185, 823. ¹H NMR (D₂O): δ (ppm) = 2.66 (dd, $J_{2a-3} = 8.53$ Hz, $J_{gem} = 16.18$ Hz, 1H, 2-H_a), 2.79 (dd, $J_{2b-3} = 5.80$ Hz, $J_{gem} = 16.18$ Hz, 1H, 2-H_b), 3.12-3.41 (m, 3H, 3-H and 4-H), 3.45 (s, 1H, C=CH), 7.28 (d, $J_{AB} = 8.23$ Hz, 2H, H_{arom}), 7.48 (d, $J_{AB} = 8.25$ Hz, 2H, H_{arom}). ¹³C NMR (D₂O): δ (ppm) = 38.8 (C-2), 40.2 (C-3), 43.9 (C-4), 79.1 (C≡CH), 83.8 (C≡CH), 121.7, 128.5, 133.3, 139.9 (C_{arom}), 175.7 (C-1). C₁₂H₁₄ClNO₂: calcd. C 60.13, H 5.89, N 5.84; found C 59.78, H 5.90 N 5.55.

2.1.15. Synthesis of (RS)-4-amino -3-(4-iodophenyl)butanoic acid hydrochloride (1c)

To a stirred solution of (*RS*)-1-*tert*-butyloxycarbonyl-4-(4-iodophenyl)-2-pyrrolidinone (**4b**) (0.25 g, 0.7 mmol) in tetrahydrofuran (6 ml) was added 1 M lithium hydroxide (2 ml). The resulting reaction mixture was stirred at ambient temperature for two hours. The solvent was evaporated under reduced pressure, the residue was diluted with water (10 ml) and extracted with diethyl ether (3 x 5 ml). The aqueous layer was acidified using 0.5 M potassium hydrogen sulfate solution, extracted with diethyl ether (3 x 15 ml). The organic extracts were dried (Na₂SO₄), and evaporated under vacuum to yield 0.23 g (81%) of (*RS*)-4-*tert*-butyloxycarbonylamino-3-(4-iodophenyl)butanoic acid (**2c**) as a pale yellow powder m.p.146-148 °C. The crude **2c** was dissolved in ~2.5 M dry hydrogen chloride/ethyl acetate solution (1.5 ml) and the reaction mixture was stirred at ambient temperature for one hour. The reaction mixture was filtered, washed with diethyl ether (5 ml). The precipitated solid was recrystallized from 2-propanol/diethyl ether to give

0.16 g, (83%) of **1c** as a white powder m.p. 208-210 °C (lit. [20] 190-195 °C). IR (neat, cm⁻¹) exhibited bands at 3154-2720, 1719, 1587, 1520, 1414, 1194. ¹H NMR (D₂O): δ (ppm) = 2.63 (dd, $J_{2a-3} = 8.55$ Hz, $J_{gem} = 16.18$ Hz, 1H, 2-H_a), 2.76 (dd, $J_{2b-3} = 5.78$ Hz, $J_{gem} = 16.18$ Hz, 1H, 2-H_b), 3.08-3.18 (m, 1H, 4-H_a), 3.24-3.36 (m, 2H, 3-H and 4-H_b), 7.04 (d, $J_{AB} = 8.55$ Hz, 2H, H_{arom}), 7.68 (d, $J_{AB} = 8.23$ Hz, 2H, H_{arom}). ¹³C NMR (D₂O): δ (ppm) = 38.4 (C-2), 39.9 (C-3), 43.9 (C-4), 93.7, 130.3, 138.5, 138.7 (C_{arom}), 175.4 (C-1). C₁₀H₁₃ClINO₂: calcd. C 35.16, H 3.84, N 4.10; found C 35.52, H 3.79 N 4.00.

2.2. Pharmacological evaluation

2.2.1. Materials

GABA was obtained from Sigma (St. Lous, MO, USA). Culture media, serum and antibiotics were obtained from Invitrogen (Paisley, UK). The rat $GABA_BR$ plasmids and the Gaqz5 construct were generous gifts from Dr. Janet Clark (National Institute of Health, Bethesda, MD, USA) and Dr. Bruce Conklin (University of California, San Fransisco, CA, USA). The tsA cells were a generous gift from Dr. Penelope S. V. Jones (University of California, San Diego, CA, USA).

2.2.2. Methods

TsA cells (a transformed human embryonic kidney (HEK) 293 cell line) [21] were maintained at 37 °C in a humidified 5% CO₂ incubator in Dulbecco's modified Eagle medium (DMEM) supplemented with penicillin (100U/ml), streptomycin (100mg/ml) and 10% fetal calf serum. One million cells were split into a 10 cm tissue culture plate and transfected the following day with 0.7 µg GABA_BR1b-pcDNA3.1, 3.5 µg GABA_BR2-pcDNA3.1 and 0.7µg Gaq-z5-pcDNA using SuperFect as a DNA carrier according to the protocol by the manufacturer (Qiagen, Hilden, Germany). The day after transfection, cells were split into one poly-D-lysine coated 96-well blackwalled-clear-bottomed tissue culture plates in the same medium as mentioned above and incubated overnight. The following day the measurement of intracellular calcium was performed as follows. The media was exchanged with Hanks balanced saline solution containing 1 mM $CaCl_2$, 1 mM MgCl_2, 20 mM HEPES, 2.5 mM probencid and 4 μ M Fluo-4AM (pH = 7.4). The cells were incubated for 1 h at 37 °C in a humidified 5% CO2 incubator. Cells were then washed twice with the same buffer without Fluo-4AM and finally 100 μ l of the buffer was left in the wells. The cell plate was then transferred to the NovoStar (BMG Labtechnologies, Offenburg, Germany) and the basal fluorescence level was adjusted to ~ 10000 fluorescence units (FU) using excitation/emission wavelengths of 485-520 nm, respectively. Fluorescence readings were measured for 45 s after addition of ligand and response was calculated as peak response minus basal level. Inactive compounds were also tested as antagonists. Twenty min after application of ligand, 10 µM GABA was added to the well and fluorescence was measured as above. All experiments were performed in triplicate and the results are given as mean \pm S.E.M of 3-4 experiments.

3. Results and discussion

3.1. Chemistry

The developed synthetic pathway for the preparation of the title compound 1a is illustrated in Scheme 1. The pathway was commenced by diazodization of 5-aminoisophthalic acid (9a) *via* adopting the procedure mentioned by Gunstone and Tucker [22]. The produced crude 5chloroisophthalic acid (8a) was subjected to esterification using the procedure described by Dimick *et al.* [23] in methanol and a catalytical amount of concentrated sulfuric acid to yield diester 7a. Subsequent saponification of 7a using one mole of 1 N sodium hydroxide solution

afforded monoester 5-chloroisophthalic acid monomethyl ester (6a) which was subjected to selective reduction of carboxylic acid functionality in the presence of ester functionality using borane-dimethyl sulfide complex to afford the alcohol 5a. Elaboration of the alcohol functionality

of **5a** to the corresponding bromo derivative **4a** was accomplished by addition of phosphorous tribromide to **5a** in solvent-free medium according to the procedure mentioned by Amrollah-Madjdabadi *et al.* [24] Compound **4a** was allowed to react with sodium azide in refluxing acetone to give compound **3a** which yielded the benzoic acid derivative **2a** upon reflux in methanolic sodium hydroxide solution. Catalytic hydrogenation of the azide functionality of **2a** in 95% 2-propanol and concentrated hydrochloric acid in the presence of PtO₂ gave the target compound **3**-aminomethyl-5-chlorobenzoic acid hydrochloride (**1a**). The crude **1a** was recrystallized from 2-propanol/diethyl ether mixture to afford **1a** in 82% yield as a white hygroscopic powder. The structure of **1a** has been established through, IR, ¹H NMR, ¹³C NMR, and microanalytical data.



Scheme 1: Synthesis of the target compound 1a.

Reagents and conditions: i) 1) NaNO₂ / H_2SO_4 / 70 °C 2) CH₃COOH / 40 °C / 0.5h 3) CuCl / conc. HCl / 80 °C / 20 min; ii) CH₃OH/H₂SO₄/reflux /24h; iii) 1 N NaOH/CH₃OH/25 °C/18h; iv) Borane-dimethyl sulfide complex/ THF/25 °C /30h; v) PBr₃/-30 ° /0.5h; vi) NaN₃/acetone/H₂O/reflux/18h; vii) 1N NaOH/CH₃OH/ reflux/2.5h; viii) H₂ / PtO₂ / 40 Psi / 95% 2-propanol /25 °C / 18h.

An examination of the literature exposed that the electronegativity of the ethynyl functionality is similar to that of the chloro functionality. In addition, the ethynyl group would be able to act as a hydrogen bond acceptor or donor in the receptor active site [25]. Accordingly, we are convinced to synthesize the *para* ethynyl analogue of baclofen, namely, (*RS*)-4-amino-3-(4-ethynylphenyl)butanoic acid hydrochloride (**1b**), as depicted in Scheme 2. Thus, (*E*)-3-(4-iodophenyl)acrylic acid methyl ester (**7b**) has been successfully achieved by adopting the general trivial procedure of Wittig reaction [26] using 4-iodobenzaldehyde (**8b**) and (methoxycarbonyl-

146

methylene)triphenylphosphorane [19]. Condensation of nitromethane with compound 7b in the presence of benzyltrimethylammonium hydroxide (Triton B, ~40% solution in methanol) was done in methanol according to the procedure described by Berthelot and his co-workers [27] to furnish the nitro ester **6b**. Selective reduction of the nitro functionality of **6b** without dehalogenation of the aromatic halogen has been established by using zinc dust in methanol/concentrated hydrochloric acid mixture at 0°C for 30 min followed by treatment of the produced amino ester with 6 N NaOH solution to give the pivotal lactam (RS)-4-(4-iodophenyl)-2pyrrolidinone (5b) in 65% yield. Compound 5b is the key intermediate in the synthetic strategy employed to achieve our targets 1b and 1c. Protection of the amide nitrogen of 5b with tertbutyloxycarbonyl group has been successfully achieved via reaction of 5b with ditertbutyldicarbonate in dichloromethane in the presence of triethylamine and 4-(dimethylamino)pyridine (DMAP) to give 4b. Subsequently, Sonogashira-Hagihara coupling [28] was conducted via addition of trimethylsilylacetylene (TMSA) to a suspension of 4b in triethylamine in the presence of a catalytical amount of bis (triphenylphosphine)palladium dichloride and cuprous iodide to produce (RS)-1-tert-butyloxycarbonyl-4-(4-trimethylsilanylethynylphenyl)-2pyrrolidinone (3b). Compound 3b was subjected to cleavage of trimethylsilyl group and lactam ring opening under mild basic conditions. The presence of tert-butyloxycarbonyl as a lactam nitrogen protecting group facilitated the lactam ring opening under mild basic conditions. Thus, 1 M lithium hydroxide solution was added to a solution of 3b in tetrahydrofuran and the reaction mixture was stirred at room temperature for 18h to afford (RS)-4-tert-butyloxycarbonylamino-3-(4-ethynylphenyl)butanoic acid (2b) in 90% yield. Finally, N-boc protecting group in compound **2b** has been cleanly removed by stirring a solution of **2b** in ~ 2.5 M dry hydrogen chloride/ethyl acetate at room temperature for one hour to yield crude (RS)-4-amino-3-(4-ethynylphenyl)butanoic acid hydrochloride (1b) which was recrystallized from 2-propanol/diethyl ether mixture to furnish the title compound 1b as a colourless crystals in 87% yield. The structure of 1b has been established through IR, ¹H NMR, ¹³C NMR, and microanalytical data.



Scheme 2: Synthesis of the target compounds 1b and 1c.

(*RS*)-4-Amino-3-(4-iodophenyl)butanoic acid hydrochloride (1c) is a baclofen analogue containing the iodo functionality as a bioisostere for the chloro functionality in the *para* position of baclofen. Compound 1c has been successfully achieved from the lactam 4b by lactam ring opening and deprotection of the *N*-boc functionality as illustrated in Scheme 2. The structure of 1c has been established through IR, ¹H NMR, ¹³C NMR, and microanalytical data.

3.2. GABA_B receptor agonistic activity

The synthesized compounds **1a-c** were evaluated for their GABA_BR agonist activity on tsA cells transfected with GABA_{B1b}/GABA_{B2}/Gaq-z5 based on Ca²⁺ measurement rather than measurement of inositol phosphate generation as previously published [29].

The GABA_B agonist activity of the synthesized compounds **1a-c** is summarized in Table 1. Compounds **1b** and **1c** are active as GABA_BR agonists (EC₅₀ value = 240 and 32 μ M, respectively, whereas compound **1a** (EC₅₀ > 300 μ M) is considered inactive as GABA_BR agonist in the GABA_BR subtype used in our assay. Compound **1a** was evaluated as GABABR antagonist at 1mM concentration against 10 μ M GABA, however, it was not effective as GABABR antagonist.

Compound No.	EC ₅₀ (µM)	$pEC50 \pm S.E.M$
1 a	>300	<3.52
1b	240	3.62 ± 0.1
1c	32	4.49 ± 0.1
GABA	0.84	6.08 ± 0.1
(RS)-baclofen	5.8	5.24 ± 0.1

Table 1: GABA_BR agonist of compounds 1a-c, GABA, and (RS)-baclofen

4. Conclusion

Synthesis and GABA_B receptor agonist activity of certain amino acids structurally related to the clinically used drug, baclofen (2), are reported. Inclusion of chlorophenyl moiety into DAVA backbone gave compound **1a** which was devoid of both agonist and antagonist activities on GABA_B receptors. Replacement of *para* chloro substituent of baclofen (2) with ethynyl group led to compound **1b** which exhibited GABA_B receptor agonist activity with $EC_{50} = 240 \mu M$. The iodo analogue of baclofen, compound **1c**, emerged as the best GABA_B receptor agonist congener in the synthesized compounds with $ED_{50} = 32 \mu M$.

Acknowledgment

The authors extend their appreciation to the Deanship of Scientific Research at King Saud University for funding this work through research group No. RGP-VPP-196.

References

- [1] M. Chebib, G. A. R. Johnston, Clin. Exp. Pharmacol. Physiol., 26, 937 (1999).
- [2] N. G. Bowery, D. R. Hill, A. L. Hudson, A. Doble, D. N. Middlemiss, J. Shaw, M. Turnbull, Nature, 283, 92 (1980).
- [3] K. Kaupmann, K. Huggel, J. Heid, P. J. Flor, S. Bischoff, S. J. Mickel, G. McMaster, C. Angst, H. Bittiger, W. Froestl, B. Bettler, Nature, 386, 239 (1997).

148

- [4] A. R. Calver, C. H. Davies, M. Pangalos, Neurosignals, 11, 299 (2002).
- [5] M. J. Robbins, A. R. Calver, A. K. Fillipov, A. Couve, S. J. Moss, M. N. Pangalos, J. Neurosci., 21, 8043 (2001).
- [6] F. Xu, G. Peng, T. Phan, U. Dilip, J. Lu C., T Chernov-Rogan, X. Zhang, K. Grindstaff, T. Annamalai, K. Koller, M. A. Gallop, D. J. Wustrow, Bioorg. & Med. Chem. Lett. 21, 6582 (2011) and references cited in.
- [7] A. Buda-Levin, F. H. E. Wojnicki, R. L. Corwin, Physiol. Behav., 86, 176 (2005).
- [8] R. N. Brogden, T. M. Speight, G. S. Avery, Drugs, 8, 1 (1974).
- [9] R. J. Tayacke, A. Lingford-Hughes, L. J. Reed, D. J. Nutt, Adv. Pharmacol., 58, 373 (2010).
- [10] I. Lidums, A. Lehmann, H. Checklin, J. Dent, R. H. Holloway, Gastroenterology, 118, 7 (2000).
- [11] K. Yomiya, N. Matsuo, S. Tomiyasu, T. Yoshimoto, T. Tamaki, T. Suzuki, M. Matoba, Am. J. Hosp. Palliat. Med., 26, 112 (2009).
- [12] M. C. Taylor, C. P. Bates, Br. J. Urol., 51, 504 (1979).
- [13] C. Herdeis, H. P. Hubmann, Tetrahedron: Asymmetry, 3, 1213 (1992).
- [14] M. Muhyaddin, P. J. Roberts, G. N. Woodruff, Br. J. Pharmacol., 77, 163 (1982).
- [15] P. Berthelot, C. Vaccher, A. Musadad, N. Flouquet, M. Debaert, M. J. Luyckx, J. Med. Chem., 30, 743 (1987).
- [16] T. Toda, Bull. Chem. Soc. Jpn., 40, 588 (1967).
- [17] E. L. Michelotti, F. Washington, R. R. Raney, N. Square, D. H. Young, US. Pat. 5254584 (1993).
- [18] O. Isler, H. Gutmann, M. Montavon, R. Rüegg, G. Ryser, P. Zeller, Helv. Chem. Acta, 40, 1242 (1957).
- [19] D. L. Boger, M. A. Patane and J. Zhou, J. Amer. Chem. Soc., 116, 8544 (1994).
- [20] Y. Wakita, M. Kojima, S. W. Schwendner, D. McConnell, R. E. Counsell, J. Labelled Compd. Radiopharm., 28, 123 (1990).
- [21] M. Chahine, P. B. Bennett, A. L. Jr. George, R. Horn, Pfluegers Arch., 427, 136 (1994).
- [22] F. D. Gunstone, S. H. Tucker, Org. Syn., Coll. Vol. 4, 160 (1963).
- [23] S. M. Dimick, S. C. Powell, S. A. McMahon, D. N. Moothoo, J. H. Naismith, E. J. Toone, J. Am. Chem. Soc., 121, 10286 (1999).
- [24] A. Amrollah-Madjdabadi, T. N. Pham, E. C. Ashby, Synthesis, 614 (1989).
- [25] (a) P. R. Wells, "Group Electronegatives" in *Progress in Physical Organic Chemistry*, A. Jr. Streitwieser and R. W. Taff (Eds.), John Wiley and Sons, NewYork, 6, pp. 111-146 (1968).
 (b) A. H. Stokes, Y. Xu, J. A. Daunais, H. Tamir, M. D. Gershon, P. butkerait, B. Kayser, J. Altman, W. Beck, K. E. Vrana, J. Neurochem., 74, 2067 (2000).
- [26] G. Wittig, G. Geissler, Liebigs Ann. Chem., 580, 44 (1953).
- [27] P. Berthelot, C. Vaccher, N. Flouquet; M. Debaert, M. Luyckx, C. Brunet, J. Med. Chem., 34, 2557 (1991).
- [28] B. Kayser, J. Altman, W. Beck, Tetrahedron, 53, 2475 (1997).
- [29] A. A. Jensen, B. E. Madsen, P. Krogsgaard-Larsen, H. Bräuner-Osborne, Eur. J. Pharmacol., 417, 177 (2001).